Protein A/G/L Magnetic Beads

CATALOG #: 6547-1

AMOUNT: 1 ml

LOT #: _____

PREPARATION: Protein A/G/L Magnetic Beads are prepared by covalently coupling Recombinant fusion Protein A/G/L (contains thirteen IgG binding domains, BV catalog # 6540) to 6% crosslinked magnetically beaded agarose. The coupling technique is optimized to give a high binding capacity for IgG. The capacity of IgG binding is generally greater than 10 mg of human IgG per ml of wet gel.

CONTENTS: Supplied as a 50% slurry in PBS with 0.02% sodium azide.

TECHNICAL SPECIFICATIONS:

Parameter	Description
Support Characteristics	Paramagnetic, spherical, 6 % cross-linked agarose
Ligand	Recombinant fusion Protein A/G/L
Particle Size	75 – 150 μm
Binding Capacity	Generally >10 mg human IgG/ml wet beads
Working Temperature	Room temperature
Storage Solution	PBS w/0.02% NaN_3
Storage Temperature	4 – 8 °C
Stability	Stable, as supplied, for at least 1 year.

FEATURES: Easy to use, high-binding capacity, non-adherent beads. Useful for immunoprecipitation and enrichment of IgG antibodies. High affinity for Fc region of IgG antibodies from a variety of species. Protein A/G/L binds to all IgG subclasses from various mammalian species, including all IgGs that bind Protein A, Protein G and Protein L individually, making it the ideal choice for purification of all kinds of polyclonal or monoclonal IgG antibodies.

FOR RESEARCH USE ONLY! Not to be used on humans.

SUGGESTED PROTOCOL:

Prepare the antibody solution by diluting the required amount of antibody in binding buffer before running the protocol.

1. Magnetic Bead Preparation (perform three times)
   a. Dispense the required amount of magnetic beads into a 1.5 ml microfuge tube.
   b. Place the tube in the magnetic rack and remove the storage solution.
   c. Add 500 µl binding buffer.
   d. Resuspend the beads.
   e. Remove the liquid
2. Antibody Capture
   a. Immediately add the antibody solution.
   b. Resuspend and mix (slow end-over-end) for at least 15 minutes.
   c. Remove the liquid.
3. Washing
   a. Add 500 µl Binding Buffer containing 0.5 M NaCl; Remove the liquid.
   b. Add 500 µl Binding Buffer; Remove the liquid.
4. Target Binding
   a. Add sample diluted in binding buffer.
   b. Incubate with slow end-over-end mixing for up to 60 minutes.
   c. Remove and collect unbound fraction.
5. Washing (perform three times)
   a. Add 500 µl wash buffer
   b. Remove liquid (save washes to troubleshoot)
6. Elution (perform three times)
   a. Add 2 volumes elution buffer (vs. bead volume).
   b. Completely resuspend beads and incubate at least 2 minutes.
   c. Remove and collect elution fraction.

RECOMMENDED BUFFER EXAMPLES:

Binding buffer: 50 mM Tris, 150 mM NaCl, pH 7.5

Wash buffer: 50 mM Tris, 150 mM NaCl, pH 7.5 (or add 1% Octylglucoside to this buffer)
(Could also try 1X PBS as both binding and wash buffer)

Elution buffer: 0.1 M -0.2 M Glycine pH 2.5-3.1 (or 0.1 M citric acid, pH 2.5-3.1 or 2.5 % Acetic Acid)

RELATED PRODUCTS:

Recombinant Protein A	Protein A Sepharose	Protein A Magnetic Beads
Recombinant Protein G	Protein G Sepharose	Protein G Magnetic Beads
Recombinant Protein L	Protein L Sepharose	Protein L Magnetic Beads
Recombinant Protein A/G	Protein A/G Sepharose	Protein A/G Magnetic Beads
Recombinant Protein A/G/L	Protein A/G/L Sepharose	Protein A/G/L Magnetic Beads
Protein A Polyclonal Antibody	Protein G-Biotin	Protein G-FITC
Protein G Polyclonal Antibody	Protein G Coated Plate	Glutathione Coated Plate
Protein L Polyclonal Antibody		

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